

# **Inorganic carbon acquisition and photorespiratory 2-phosphoglycolate metabolism in cyanobacteria**



**Marion Eisenhut, Martin Hagemann**

**University Rostock  
Department Plant Physiology  
Germany**

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# Inorganic carbon evolved as limiting factor

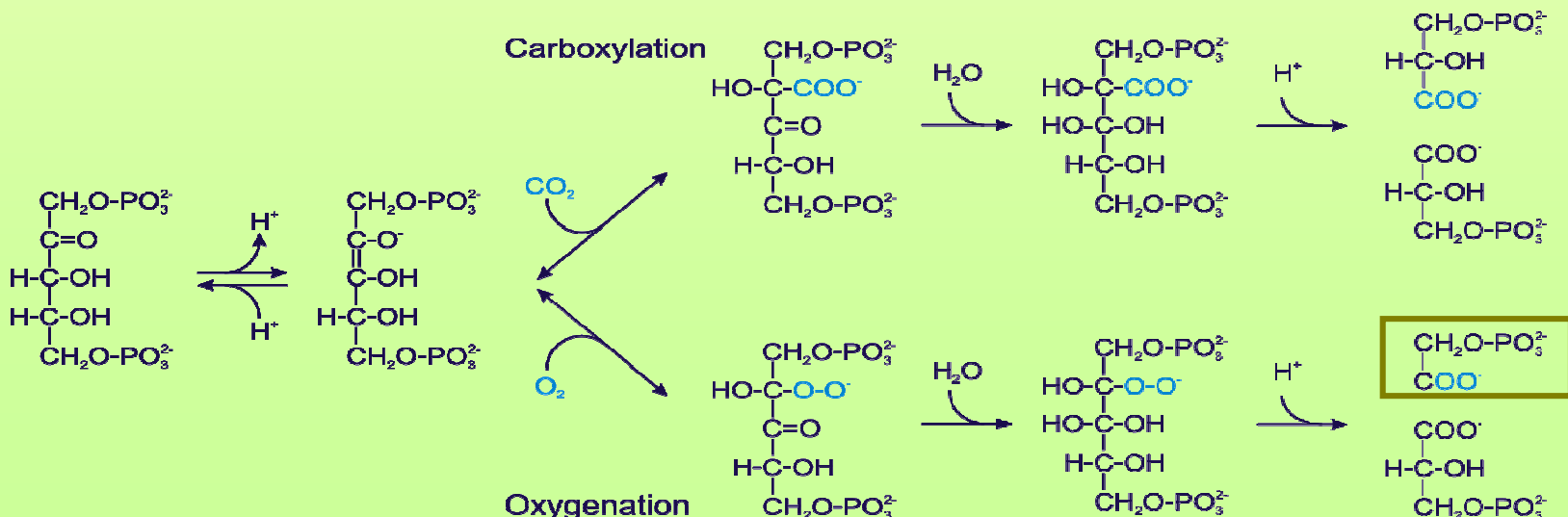
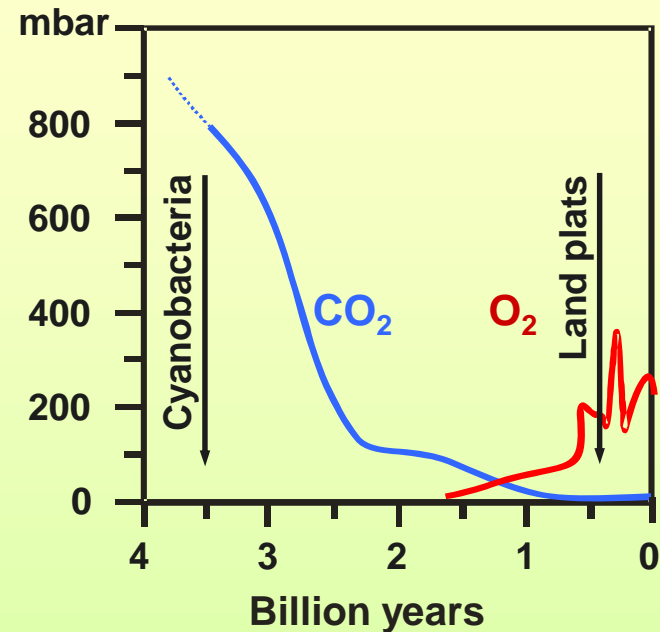
## Rubisco: an enzyme with low affinity and specificity to CO<sub>2</sub>

### 1. Problem: low affinity for CO<sub>2</sub>

	Km (CO <sub>2</sub> ) [μM]	Reference
<i>Spinacea oleracea</i>	21 ± 1	Spreitzer et al. 2005
<i>Anabaena variabilis</i>	293 ± 27	Badger 1980

→ high content of Rubisco compensates in higher plants (up to 30% leaf protein)

### 2. Problem: O<sub>2</sub> as competitive substrate



# Phosphoglycolate (2-PG) or photorespiratory cycle in plants



**Metabolic pathway transforming 2-phosphoglycolate produced by oxygenase reaction of RubisCO to 3-phosphoglycerate**

## **Functions:**

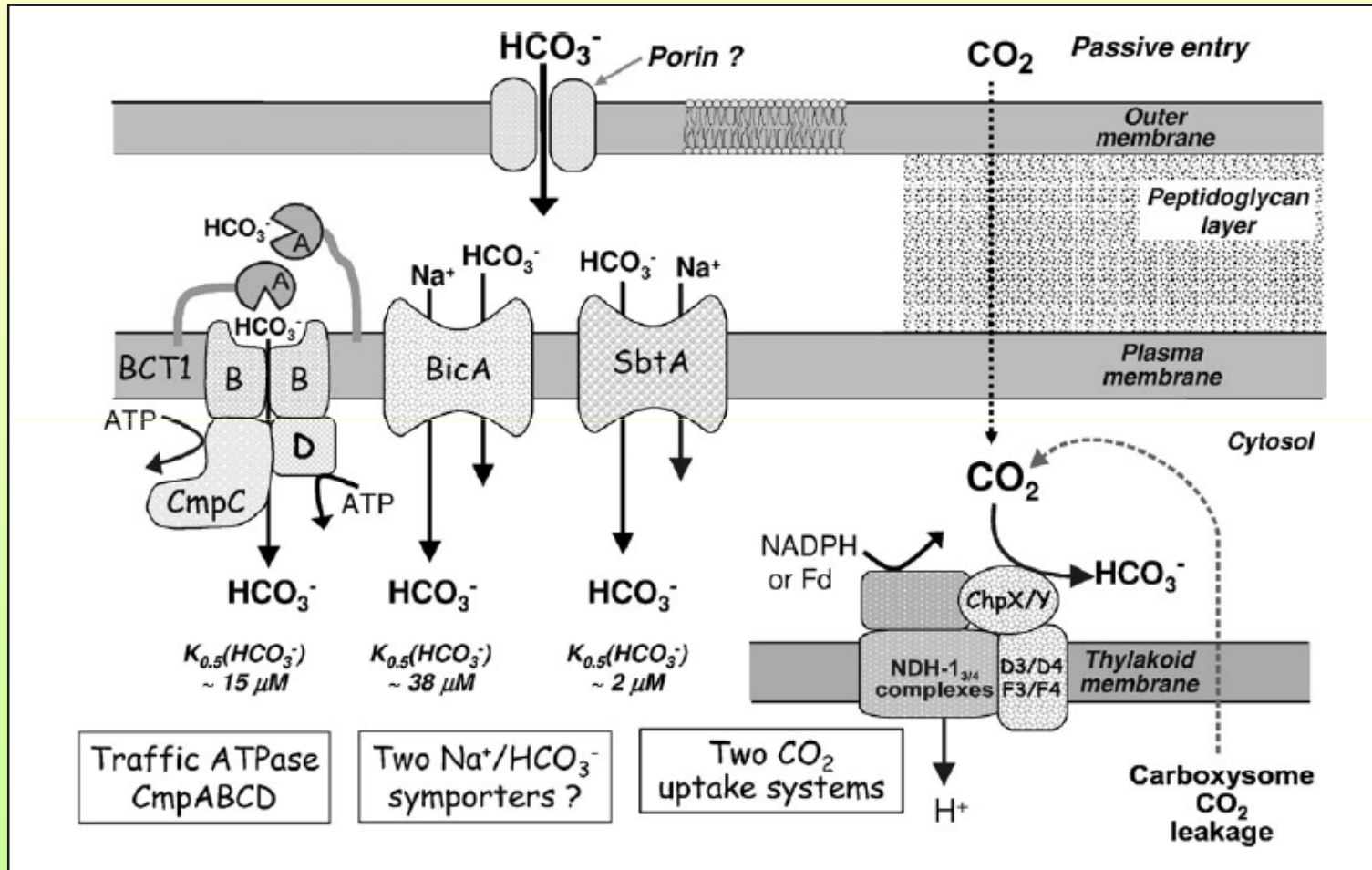
- Recycling of 75% of organic carbon from 2-PG
- Avoidance of accumulation of toxic intermediates
- Synthesis of intermediates
- Protection against high light

## **Problems:**

- Loss of organic carbon
- Loss of energy

**Photorespiratory plant mutants need high CO<sub>2</sub> – high CO<sub>2</sub>-requiring phenotype**

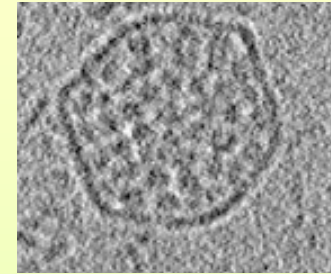
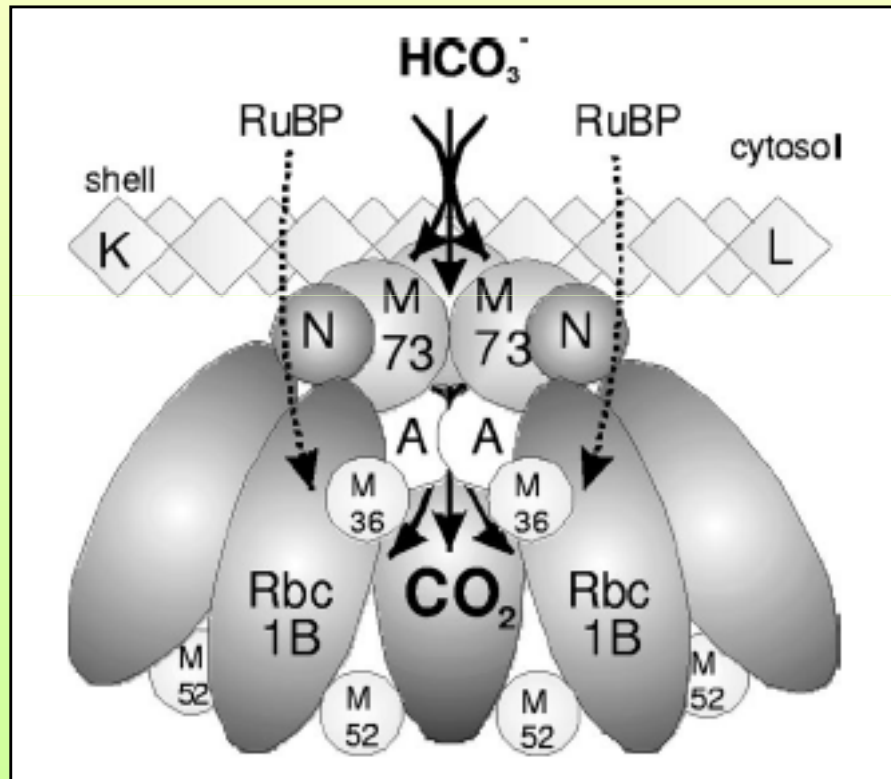
# Cyanobacteria evolved a carbon concentrating mechanism (CCM) employing C<sub>i</sub>-uptake mechanisms and carboxysomes



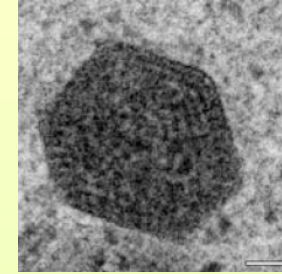
➔ Bicarbonate is concentrated inside the cell

# Cyanobacteria evolved a carbon concentrating mechanism (CCM) employing $C_i$ -uptake mechanisms and carboxysomes

Model of the bicarbonate dehydrating complex inside the carboxysome (Cot, So & Espie, 2008)



Iancu et al. 2007



Kerfeld et al. 2005

Rubisco (Rbc) is concentrated in **carboxysomes**, where carbonic anhydrase (CcaA) releases high  $CO_2$  amounts from  $HCO_3^-$

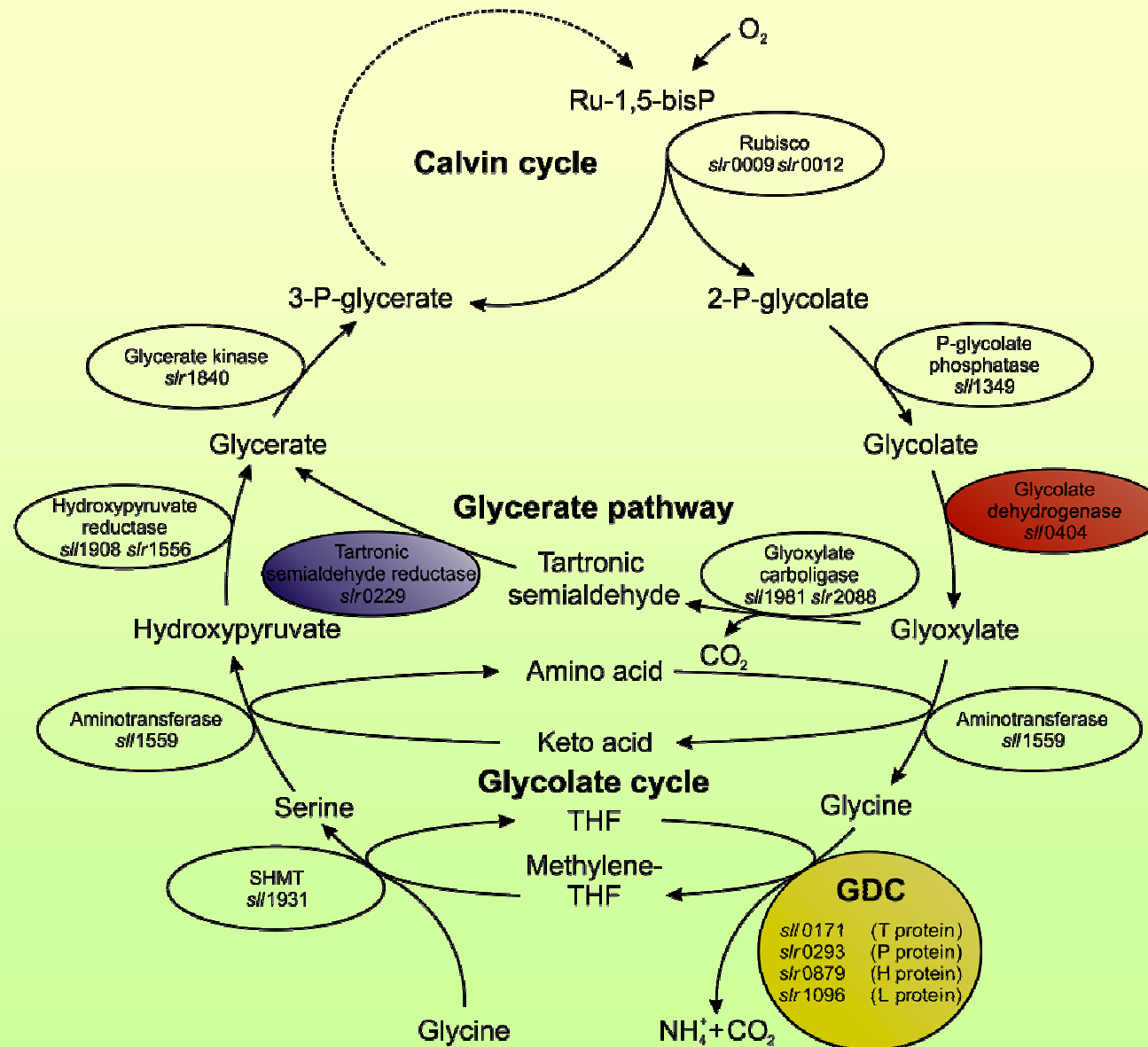
➔  $CO_2$  is released near RubisCO allowing efficient carboxylation, CCM mutants need high  $CO_2$  – high  $CO_2$ -requiring phenotype

# Search for 2-PG metabolizing proteins in *Synechocystis*: A mixture of plant and bacterial enzymes was found!

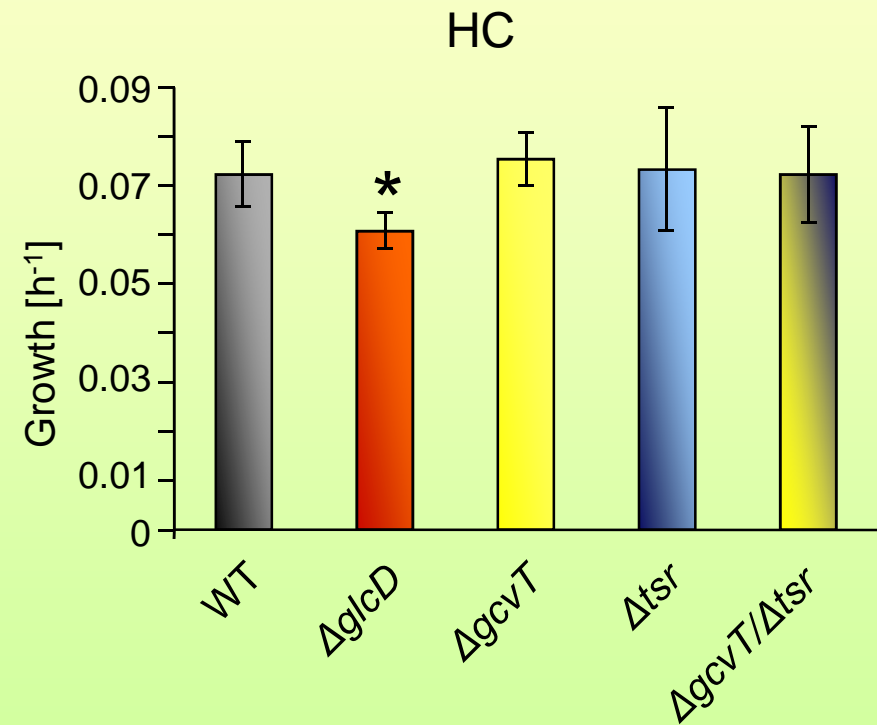
Sequence comparison of proteins participating in phosphoglycolate turnover with candidate proteins from *Synechocystis* sp. strain PCC 6803 using PSI- and PHI-BLAST (Altschul et al., 1997). The similar proteins from bacteria or *Arabidopsis thaliana* are in most cases biochemically characterized.

Protein (Abbreviation)	Organism	Acc. No.	Literature	Similarity e-value	ORF in <i>Synechocystis</i>
Phosphoglycolate phosphatase (PGP)	<i>A. eutrophus</i>	P40852	Schäferjohann et al., 1993	$3e^{-08}$	<i>slr0458</i>
				$9e^{-06}$	<i>slr1349</i>
Glycolate dehydrogenase subunit D (GlcD)	<i>E. coli</i>	AAC76015	Pellicer et al., 1996	$3e^{-126}$	<i>slr0404</i>
Serine:glyoxylate aminotransferase (AGT)	<i>A. thaliana</i>	At2g13360	Liepman and Olsen, 2001	$3e^{-59}$	<i>slr1559</i>
GDC, P protein (GcvP)	<i>A. thaliana</i>	At2g26080	Bauwe et al., unpubl.	0	<i>slr0293</i>
GDC, T protein (GcvT)	<i>A. thaliana</i>	At1g11860	Bauwe et al., unpubl.	$7e^{-52}$	<i>slr0171</i>
GDC, H protein (GcvH)	<i>A. thaliana</i>	At2g35120	Bauwe et al., unpubl.	$2e^{-28}$	<i>slr0879</i>
GDC, L protein (GcvL)	<i>A. thaliana</i>	At3g16950	Bauwe et al., unpubl.	$1e^{-164}$	<i>slr1096</i>
Serine hydroxymethyltransferase (SHMT)	<i>A. thaliana</i>	At4g37930	Voll et al., 2006	$6e^{-94}$	<i>slr1931</i>
Hydroxypyruvate reductase (HPR)	<i>A. thaliana</i>	At1g68010	Bauwe et al., unpubl.	$2e^{-28}$	<i>slr1908</i>
				$2e^{-26}$	<i>slr1556</i>
Glycerate kinase (GLYK)	<i>E. coli</i>	AAB93855	Cusa et al., 1999	$3e^{-64}$	<i>slr1840</i>
Glyoxylate carboligase (GCL)	<i>E. coli</i>	AAA23864	Chang et al., 1993	$1e^{-94}$	<i>slr2088</i>
				$3e^{-58}$	<i>slr1981</i>
Tartronic semialdehyde reductase (TSR)	<i>E. coli</i>	P77161	Cusa et al., 1999	$3e^{-35}$	<i>slr0229</i>

# Cyanobacterial 2-PG metabolism: A combination of plant-like 2-PG cycle and bacterial-like glycerate pathway?

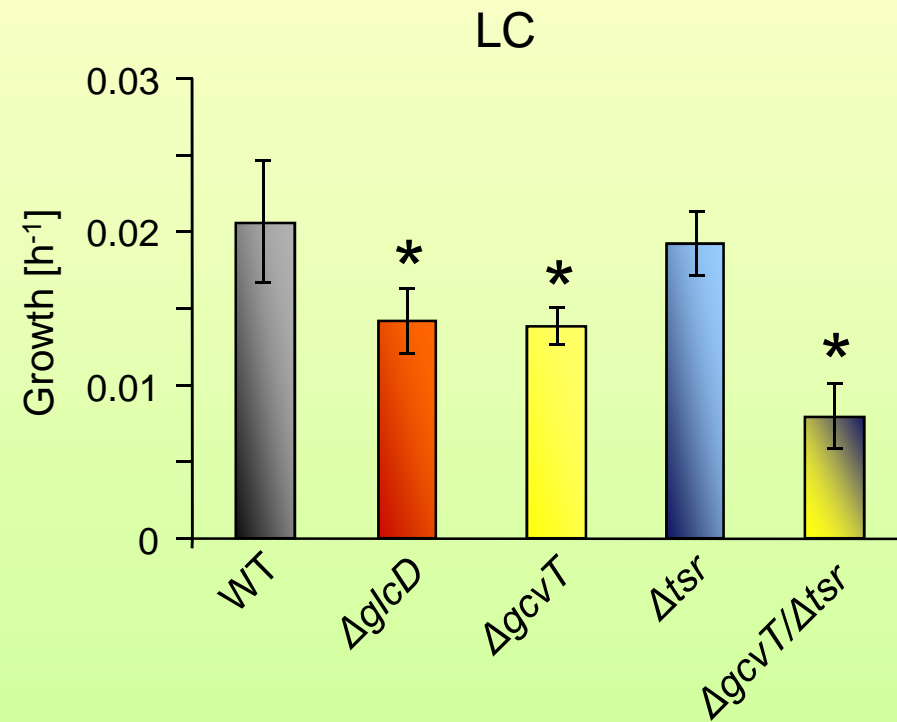


# The glycolate dehydrogenase mutant showed diminished growth at HC, while other mutants grew like WT

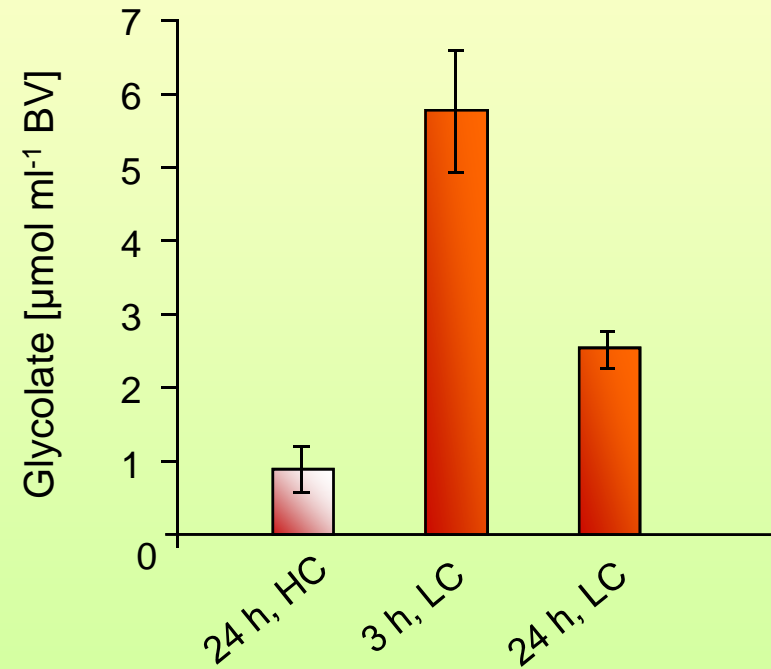




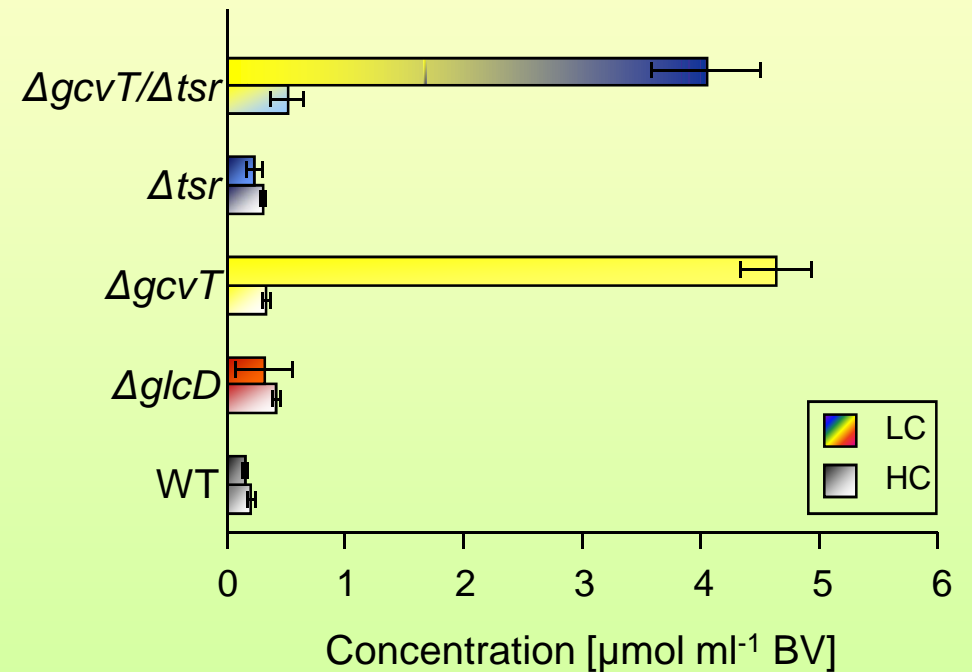
# Mutants defective in plant-like 2-PG cycle showed diminished growth at LC, while glycerate cycle mutant grew like WT



**Glycolate accumulation was only observed in cells of the glycolate dehydrogenase mutant, even at HC!**



# Glycine accumulation in mutants of ORFs encoding plant-like 2-PG cycle or/and bacterial-like glycerate pathway proteins



➔ Glycolate metabolism exists but employs at least two pathways in cyanobacteria