

Innovative Technologies for Cryopreservation of Cancer-Relevant Samples for Biobanking

BIOBANKS
Sant Feliu, 1-7 Nov 2008

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BioBank

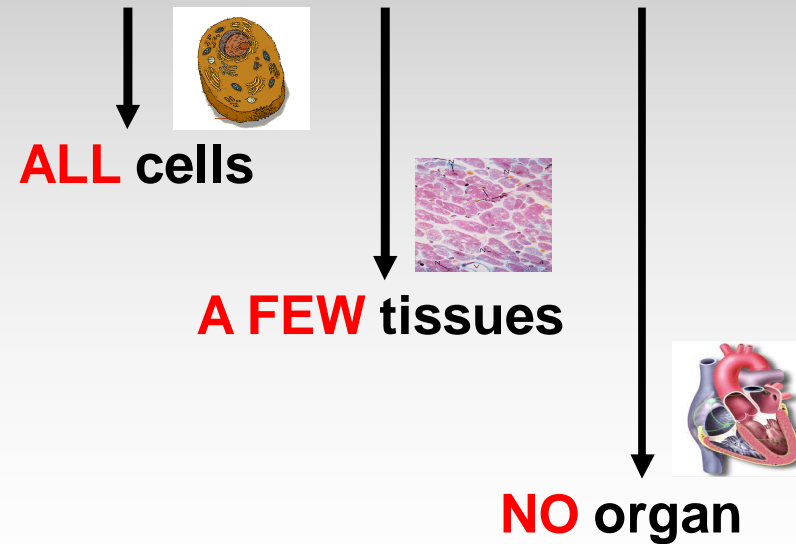
βίος = Life Bank = Store

Store of Life

Most of tissues in BioBanks are
DEAD!

CryoPreservation

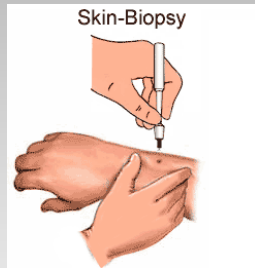
Preservation of
Life
by Cold



Current Biopsy Preservation VS. Preservation of Functionality

Preservation of Functionality

Biopsy
↓
Quenching in LN2



- AP
- DNA Sequencing
- Immunoassays



• MOLECULAR PATHWAYS

• GENE EXPRESSION

• CORRELATION WITH
SECONDARY TUMORS

• CIRCULATING TUMOR
CELLS (CTC)

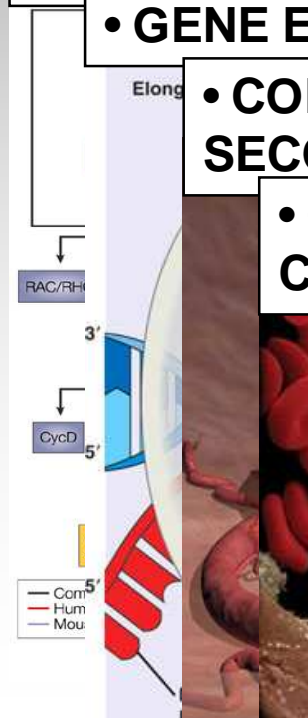
• DRUGS INTERFERENCE

• PROTEOMICS

• DIAGNOSIS

• PROGNOSIS

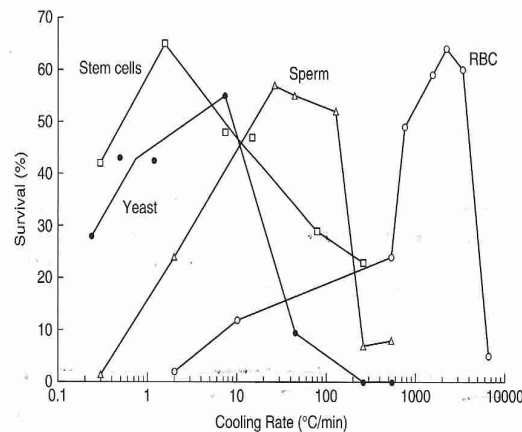
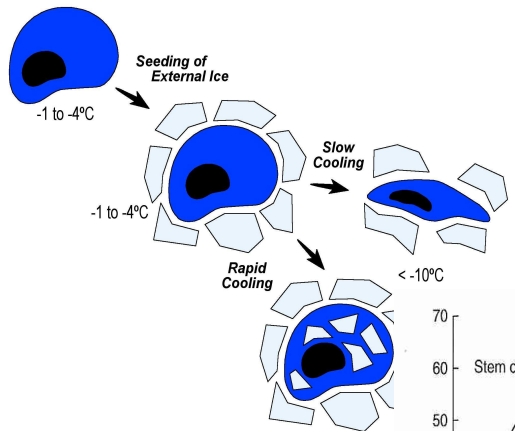
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Conventional Cryopreservation

Slow Freezing



DRAWBACKS

- Isolated cells in suspension
- Seeding of ice
- Controlling cooling rate
- No tissues
- Osmotic and Mechanical Stress
- Low recovery rates in some cell types

Vitrification

24

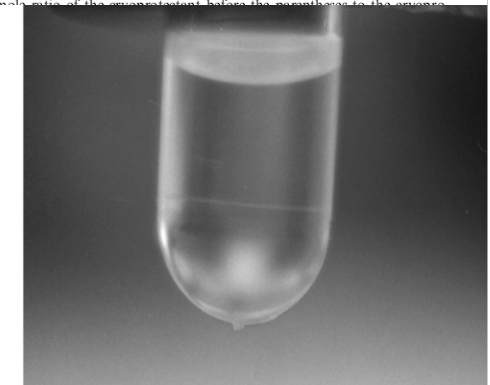
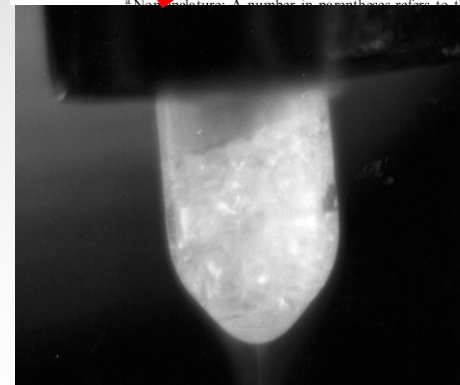
G.M. Fahy et al. / Cryobiology 48 (2004) 22

Table 1
Composition and properties of 15 model vitrification solutions

Solution number	Solution name ^a	Total % w/v	Gram quantities per ml					PEG (grams/ml)	Density (g/ml)	K ⁺ /Na ⁺ ^c			
			D	F	E	P	S						
1	D+PVP	47	41	0	0	6	0	1.0775	6.02	0.85 ± 0.06			
2	D(2)FP ₁₀ +PVP	49	36.62	0.38	0	12	0	1.0718	3.28	3.49 ± 0.22			
3	D(1)FP ₁₀ +PVP	50.5	38.88	0	0	11.62	0	1.0716	2.66	4.13 ± 0.15			
4	D(8)FP ₁₀ +PVP	51	39.34	1.66	0	10	0	1.0749	2.49	4.22 ± 0.21			
5	D(1)AP ₁₀ +PEG	48	37.08	0	10.92	0	10	0	6	1.0592	3.39	3.51 ± 0.20	
6	D(8)AP ₁₀ +PEG	49	37.19	0	10.31	0	10	0	6	1.0605	3.07	3.31 ± 0.18	
7	D(3)EP ₁₀ +PEG	49	37.08	0	10	0	6.91	10	0	6	1.0703	3.67	2.07 ± 0.12
8	D(8)EP ₁₀ +PEG	49	37.39	0	0	14.61	10	0	0	6	1.0701	3.10	2.89 ± 0.13
9	D(8)EP ₁₀ +PEG	48	35.06	0	0	15.94	10	0	0	6	1.0698	3.12	3.30 ± 0.12
10	D(8)EP ₁₀ +PEG	48	35.50	0	0	21.00	10.50	6	0	1.0702	2.82	3.41 ± 0.11	
11	D(8)EP ₁₀ +PEG	50	0	0	0	44.00	0	6	0	1.0770	2.11	3.06 ± 0.08	
12	D(8)EP ₁₀ +PEG	42	0	0	0	0	36.00	6	0	1.0503	3.47	2.96 ± 0.13	
13	D(8)EP ₁₀ +PEG	46	20.00	0	0	0	20.00	6	0	1.0640	4.02	2.34 ± 0.13	
14	D(8)EP ₁₀ +PEG	46	0	0	0	20.00	20.00	6	0	1.0619	2.67	2.92 ± 0.08	
15	D(1)A ₁₀ +PVP	45	11.10	0	8.40	0	19.50	6	0	1.0572	3.36	3.05 ± 0.16	

^a These solutions vitrify upon cooling at about 10°C/min at 100 MPa [18]. The C_v of solution 6 was interpolated. For solution 3, the estimated C_v was higher than the actual C_v by 1.5% w/v permeating cryoprotectants.

^b Note: A number in parentheses refers to the molar ratio of the cryoprotectant before the cryoprotectant to the cryoprotectant.



Our Technologies

Quartz Capillaries & Slush

SLOW FREEZING

Advantages

- Low cpa concentration
- Low toxicity

Disadvantages

- Control Cooling rate
- Need of Seeding
- Long exposition to cpa
- Very cell-type dependent
- Expensive equipment

VITRIFICATION

Advantages

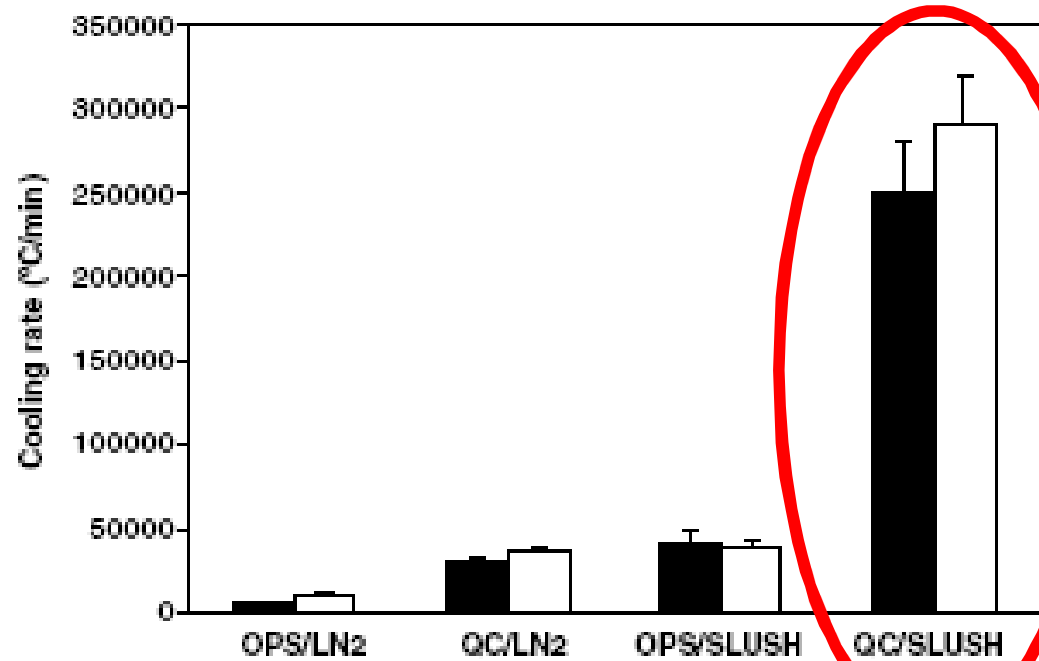
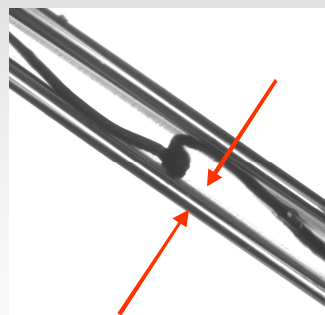
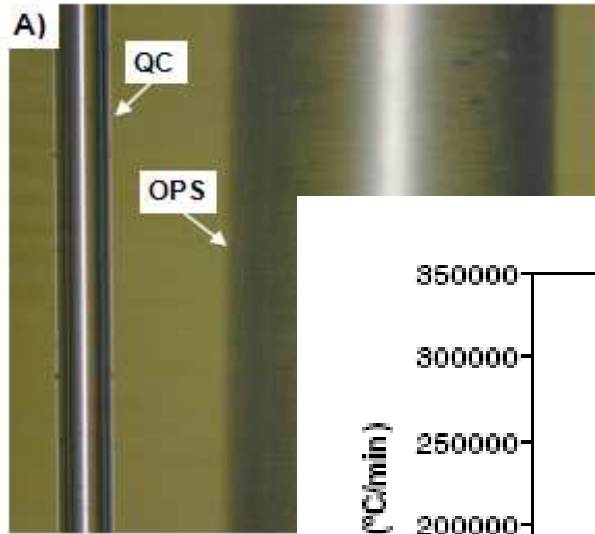
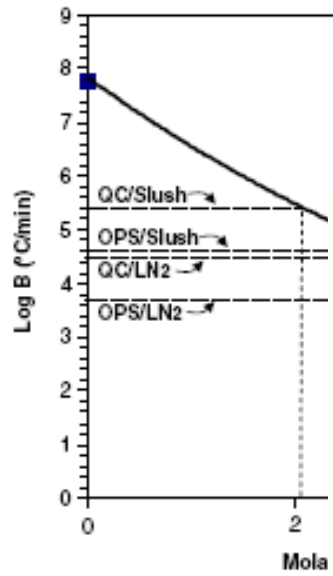
- No control of cooling rate
- No seeding
- Low sensitive to the cell type
- Cheap equipment

Disadvantages

- High cpa concentration
- High toxicity

Equilibrium Vitrification

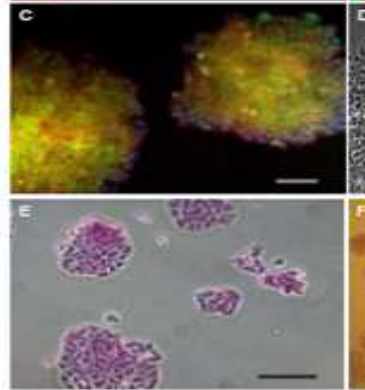
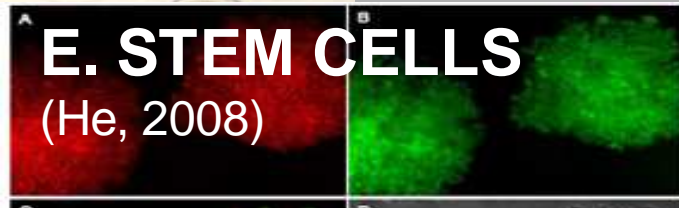
Quartz Capillaries & Slush



Source: Thermal performance of quartz capillaries for vitrification. Ramon Risco et al. Cryobiology 55 (2007) 222–229

Quartz Capillaries & Slush

Successful Applications:



C. ELEGANS

(Risco, 2008)



Equilibrium Vitrification

1284

MECHANISM OF CELL I

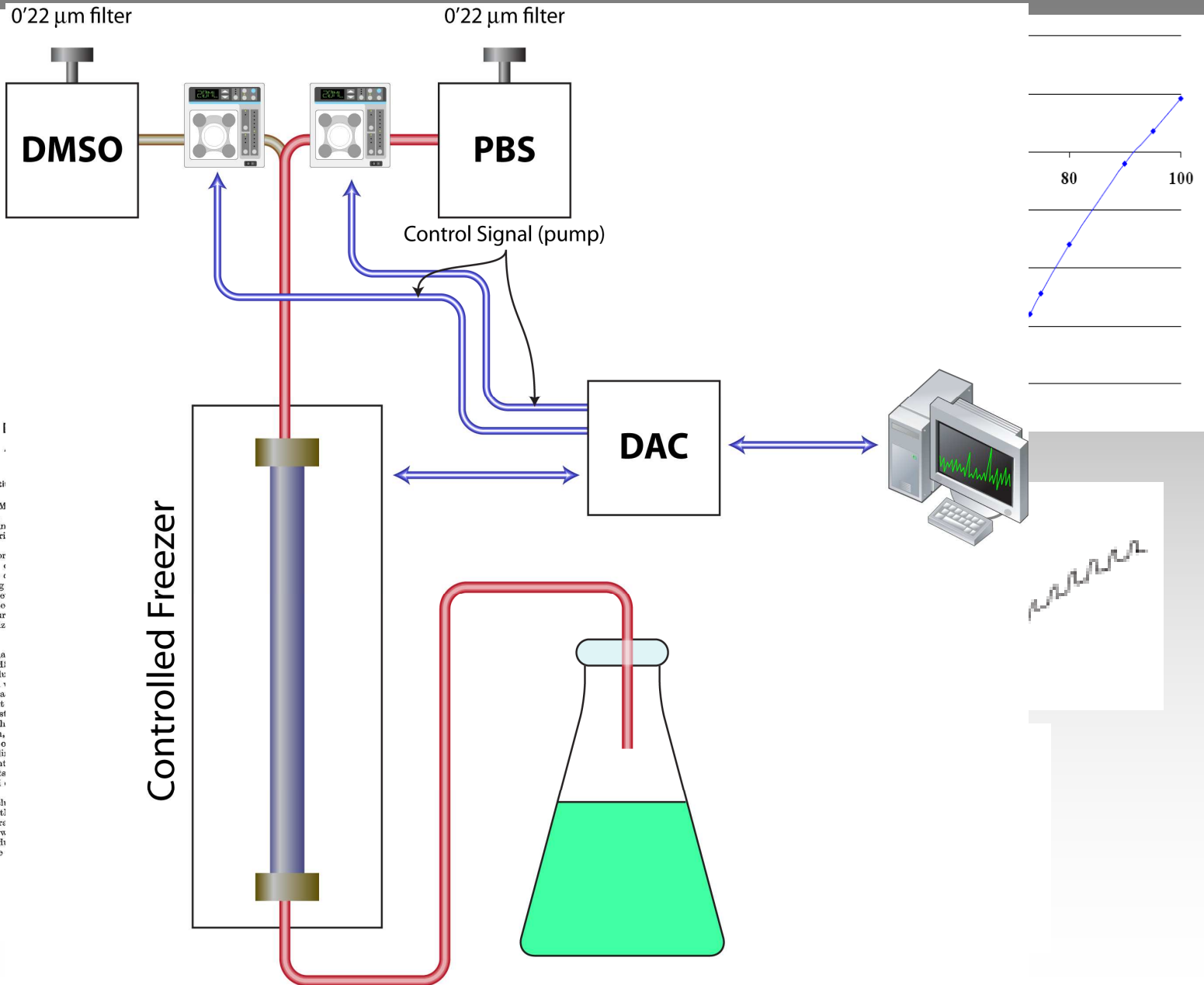
National Insti

GLYCEROL, dimethyl sulphoxide (DMSO) and other hydrophilic non-electrolytes, many types from damage during freezing and allow them to be stored for long periods.

Theories put forward to explain the prevention of crystal formation, and reduction of the concentration of electrolytes at any temperature during freezing support the latter. Two methods have been developed to prevent any rise in electrolyte concentration during cooling to low temperature: the functional recovery of an organism after re-warming has been greatly improved.

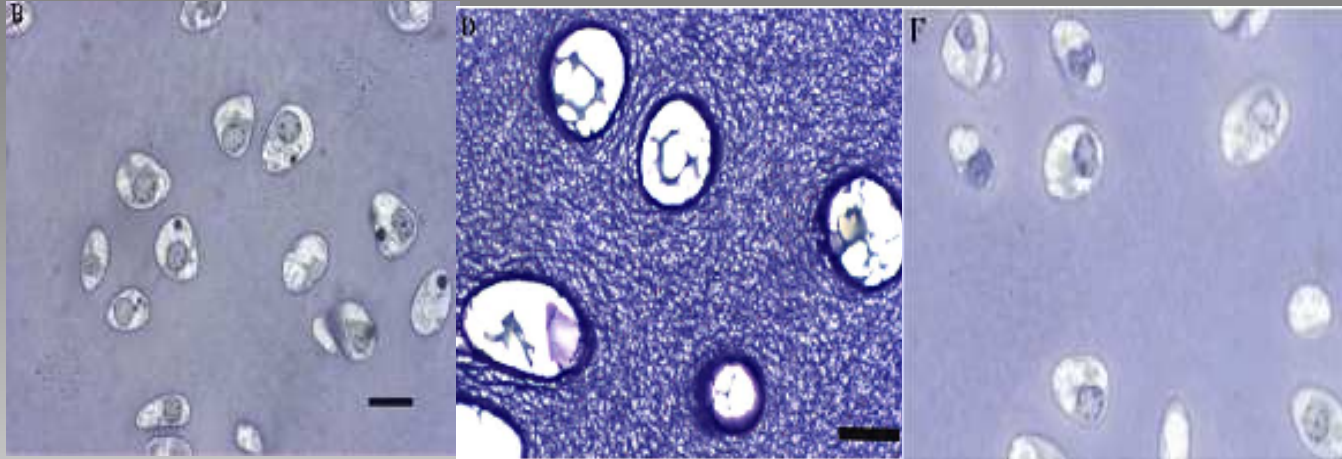
The basis of these methods is as follows. Fig. 1A shows a solid three-dimensional diagram of the three-component system DMSO-NaCl-water. The letter *a* in Fig. 1B denotes a solution of 0.154 M NaCl in a mixture of DMSO and water. Its freezing-point is at *b*. It is clear that on cooling (*bcd*) ice only will separate out and the concentrations of both the NaCl and the liquid phase increase. For comparison, the cooling of the two-component system of NaCl in water, and *ijkl* indicates the cooling of the three-component system of DMSO (10 per cent).

The curves in Fig. 2 show the effects of different initial DMSO concentrations on the NaCl concentration during freezing. The concentration of NaCl is 0.154 M. The inclusion of as little as 5 per cent v/v DMSO greatly reduces the concentration of NaCl at any temperature as ice separates. The concentration of DMSO not only does not fall but also increases progressively, but also markedly reduces the concentration of salt as the temperature

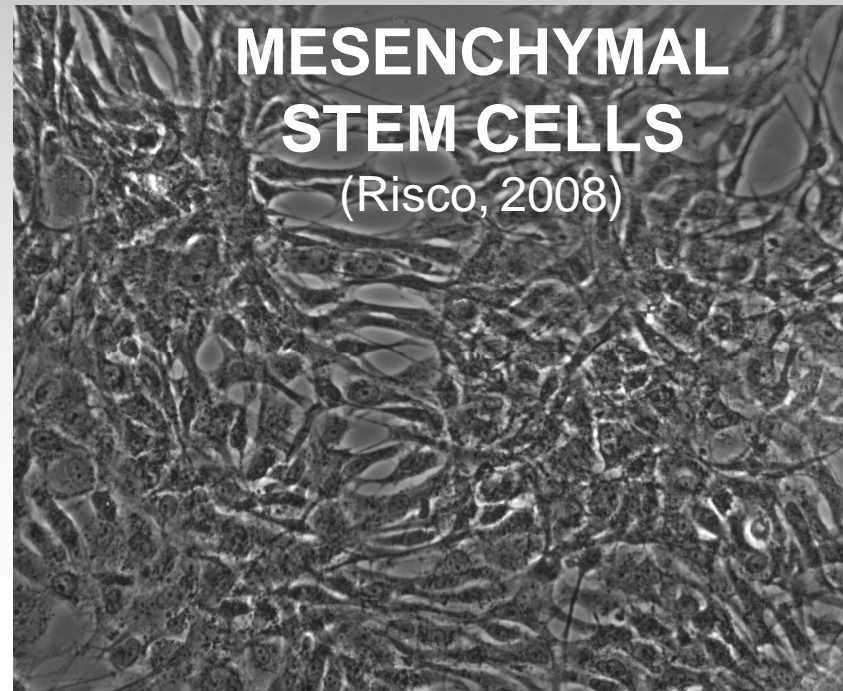
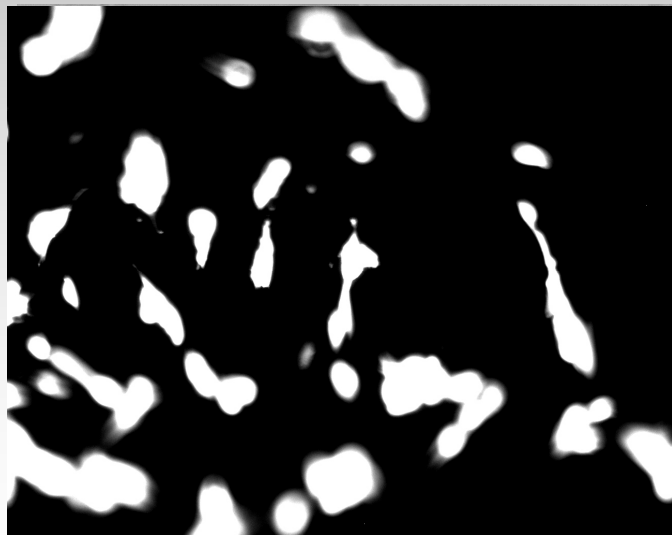


Equilibrium Vitrification

A. CARTILAGE



86.7% \pm 1.8%
incorporation
of ^{35}S sulfate
into GAGs
(Pegg, 2007)

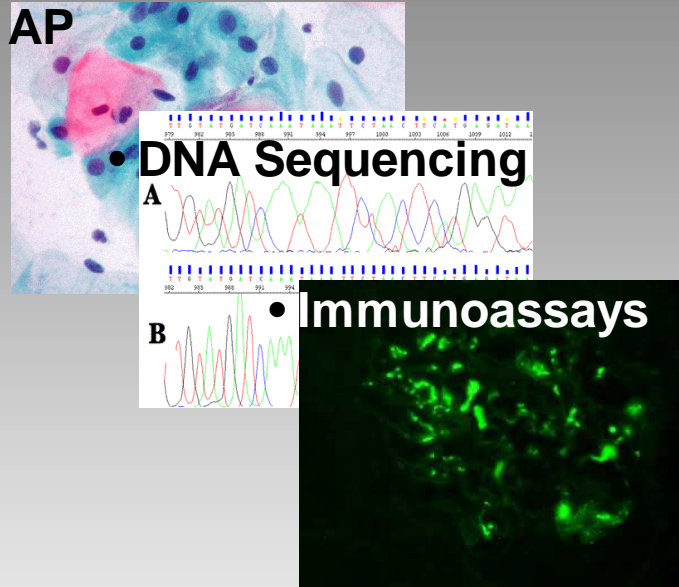


CONCLUSIONS

PRESENT STATUS OF BIOBANKING

AP

- DNA Sequencing
- Immunoassays



Collaborators:

- Mehmet Toner (Harvard Uni.)
- Alberto Olmo (Univ. Seville)

CAPABILITIES OF NOWADAYS CRYOPRESERVATION TECHNOLOGIES

• MOLECULAR PATHWAYS

• GENE EXPRESSION

• CORRELATION WITH SECONDARY TUMORS

• CIRCULATING TUMOR CELLS (CTC)

• DRUGS INTERFERENCE

• PROTEOMICS

• DIAGNOSIS

• PROGNOSIS

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