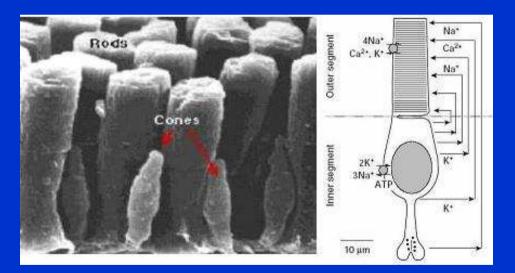
ESF-FWF Conference in Partnership with LFUI Water Interfaces in Physics, Chemistry and Biology: A Multi-Disciplinary Approach Universitaetszentrum Obergurgl (Oetz Valley, near Innsbruck), Austria 8-13 December 2007

- Hydration in GPCR-mediated signal transduction
- Joseph Parello¹, Lixin Shen¹, Jean-Louis Banères² & Michel Laguerre³
- ¹ Department of Pharmacology, Vanderbilt University School of Medicine, Nashville, TN, USA
- ² Institut des Biomolécules Max Mousseron (IBMM), UMR 5247 CNRS, Montpellier, France
- ³ Institut Européen de Chimie et de Biologie (IECB), UMR 5248 CNRS CBMN, Pessac, France.
- Abbreviation : G-protein coupled receptor (GPCR)



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Structural organization of a salamander rod

Micrograph of salamander retina

Rhodopsin is a transmembrane G protein coupled receptor (GPCR) involved in the vision process of a variety of organisms



Advances in Protein Chemistry

Volume 74, 2007, Pages 67-93 Mechanisms and Pathways of Heterotrimeric G Protein Signaling How do Receptors Activate G Proteins? William M. Oldham and Heidi E. Hamm

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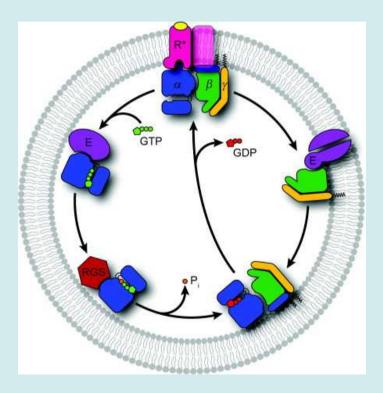


Fig. 1. Overview of the G protein cycle

Heterotrimeric G proteins GDP-bound α (*blue*), β (*green*), and γ (*gold*) subunits.

Heptahelical receptor (pink) in the cell membrane

Downstream effector proteins (E, *purple*).

Signal terminated on the hydrolysis of GTP to GDP by $G\alpha$, which may be catalyzed by RGS proteins (*dark red*).

The concept of an heptahelical organization for the transmembrane G protein coupled receptors rests upon the initial structural determination of the structure of bacteriorhodopsin (bRho)

Three-dimensional model of purple membrane obtained by electron microscopy R.Henderson & P.N.T.Unwin, Nature 1975, 257, 28-32.

A 7-A resolution map of the purple membrane has been obtained by electron microscopy of tilted, unstained specimens. The protein in the membrane contains seven, closely packed, alpha-helical segments which extend roughly perpendicular to the plane of the membrane for most of its width.



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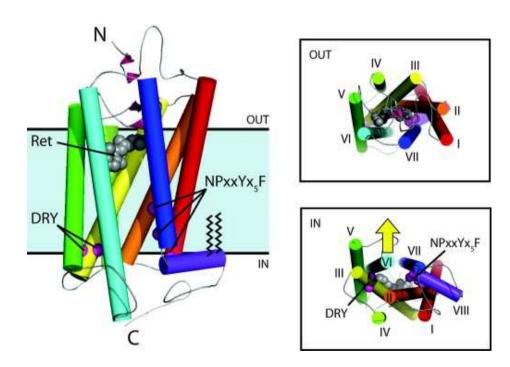
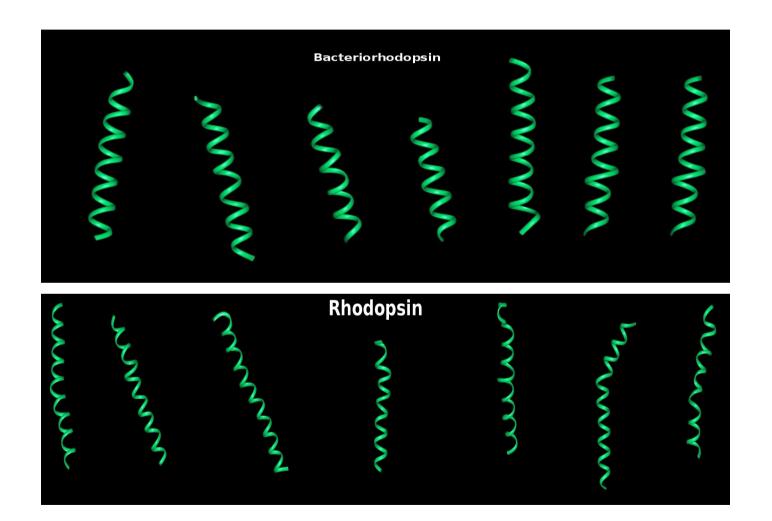


Fig. 2. Structure of a heptahelical receptor.

Cartoon model of dark (inactive) bovine rhodopsin (1U19), showing the seven transmembrane-spanning α helices (*red* to *blue*) and 11-*cis*-retinal (*gray spheres*).

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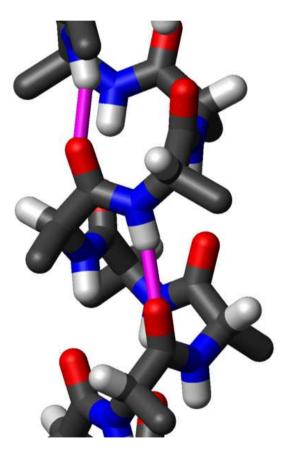


- Crystal structure of the unique parvalbumin component from muscle of the leopard shark (Triakis semifasciata). The first X-ray study of an alpha-parvalbumin.
 - Roquet F; Declercq J P; Tinant B; Rambaud J; Parello J J Mol Biol 1992, 223, 705-20.

Introduction of the plots distance (Oi/Ni+3, Oi/Ni+4) versus residue i to visualize the local geometry of alpha-helices in the protein 3D-structures

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<u>Alpha helix</u>

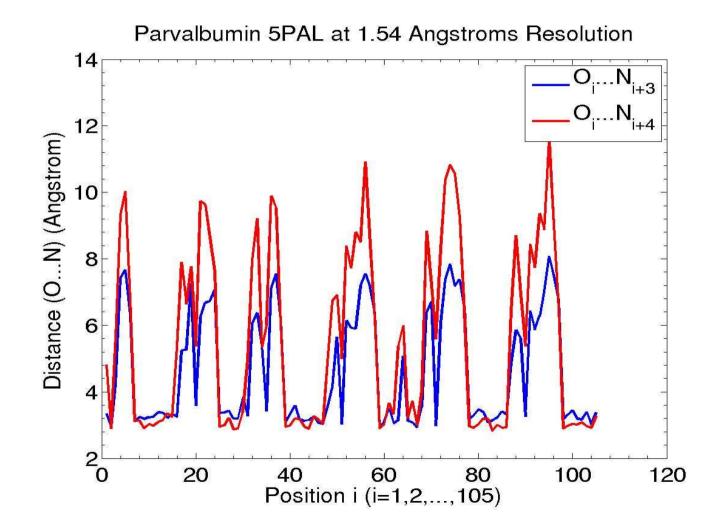
Close-up sideview of a "stick" model of an alpha helix of poly-alanine using the dihedral angles φ =-60° and ψ =-45° and the Engh&Huber bond geometry. Two hydrogen bonds are highlighted in magenta; the O-H distance is 2.08 Å.

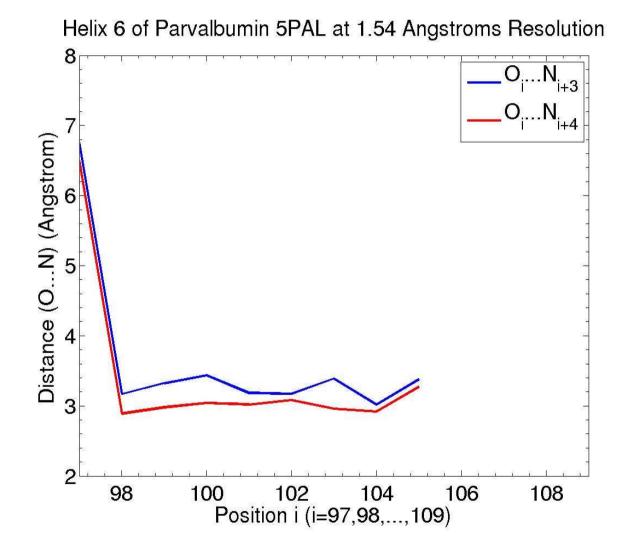
From Wikipedia, the free encyclopedia

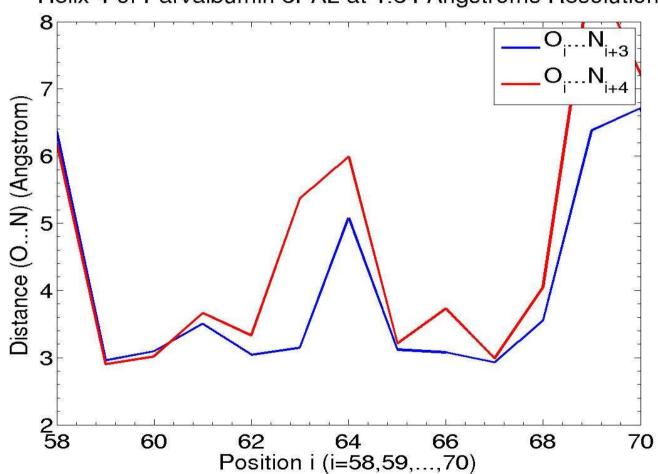
The Oi ...Ni+3 and Oi...Ni+4 distances are "magic" distances since they bring together a C=O group (i residue) and two NH groups (i+3 and i+4 residues) at hydrogen bond distance.

In the regular alpha-helix we have :

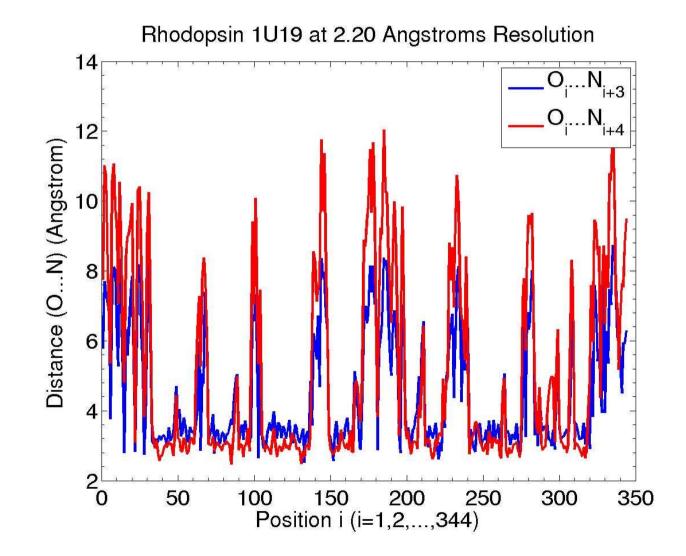
distance(Oi ...Ni+3) > distance(Oi...Ni+4)

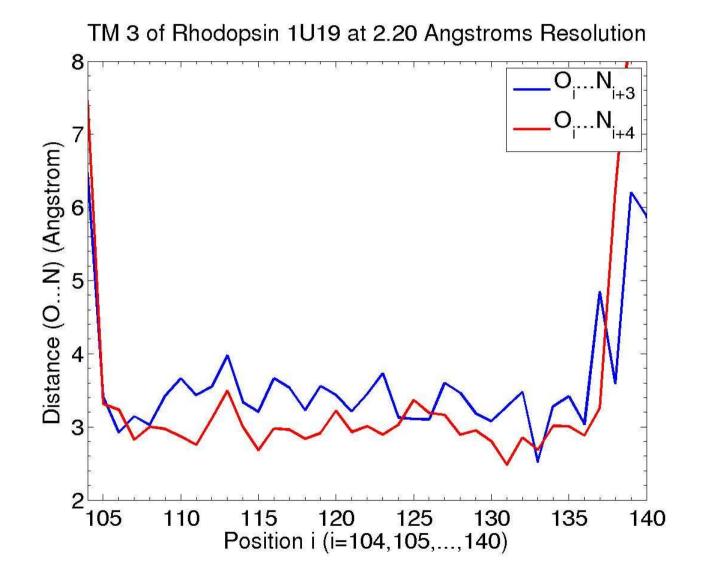


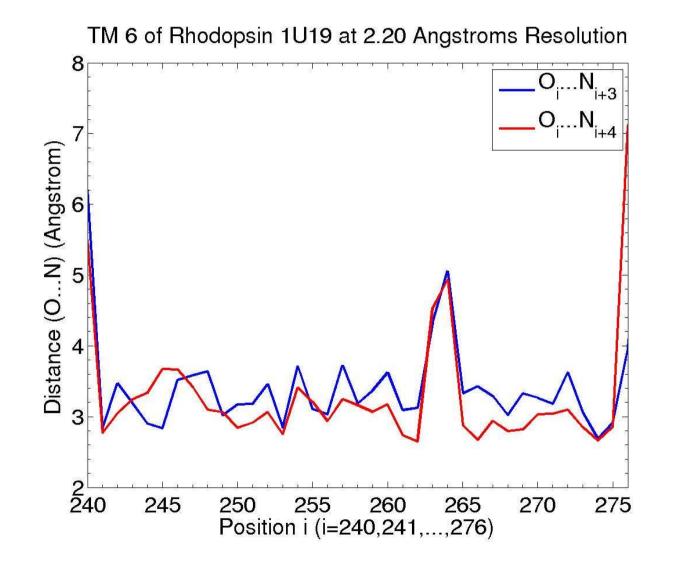


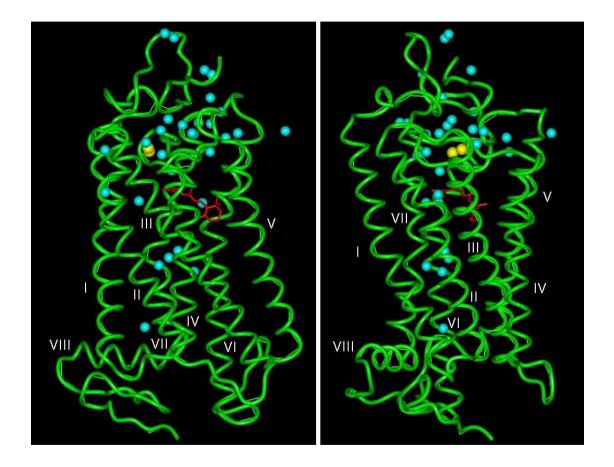


Helix 4 of Parvalbumin 5PAL at 1.54 Angstroms Resolution

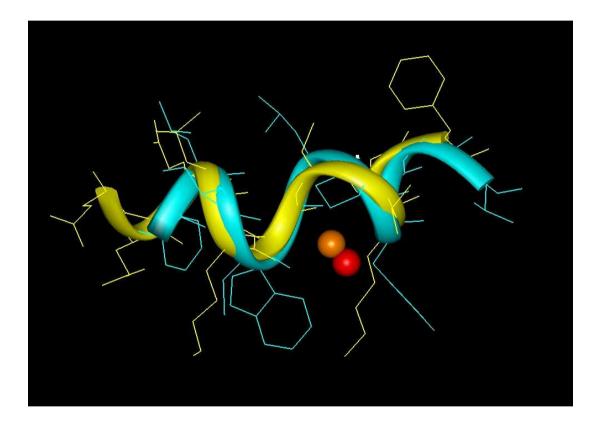








Hydration of bovine rhodopsin in the crystal structure 1U19



Selective hydration of two kinked alpha-helices with highly related geometries Blue: helix TM6 of bovine rhodopsin 1U19 Yellow: helix 4 (or helix D) of parvalbumin 5PAL Structure-based Analysis of GPCR Function: Conformational Adaptation of both Agonist and Receptor upon Leukotriene B4 Binding to Recombinant BLT1

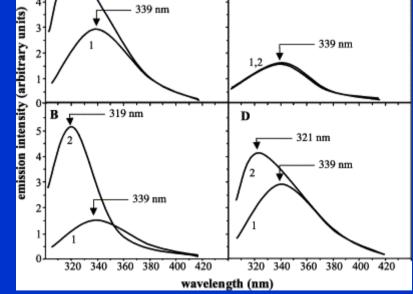
Jean-Louis Baneres, , Aimée Martin, Pierre Hullot, Jean-Pierre Girard, Jean-Claude Rossi and Joseph Parello, J Mol Biol 2003,329, 801-814

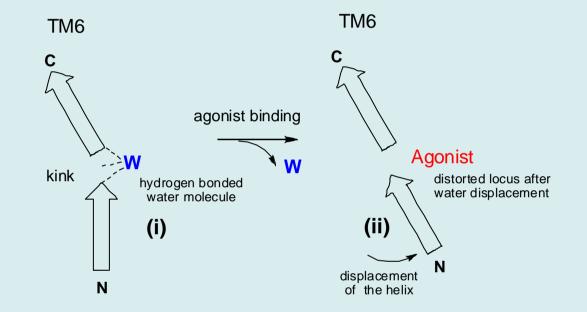
Activation of a rhodopsin-like GPCR by its natural agonist

All Trp included wt receptor

(profiles 1: free receptor; profiles 2: agonist-loaded receptor)

All Trp mutated for Leu except Trp in TM6 (kinked region) All Trp conserved except Trp in TM6 mutated for Leu (kinked region)





Hypothetical model showing the role of the central water (W) molecule at the level of the kinked region of TM6 in a GPCR: schematic presentation of the helix in the absence of agonist in the presence of water (i) and after agonist binding upon water displacement (ii)

Acknowledgements

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- Institut des Biomolécules Max Mousseron (IBMM), UMR 5247 CNRS, Montpellier, France
- Institut Européen de Chimie et de Biologie (IECB), UMR 5248 CNRS CBMN, Pessac, France
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